

# Relationship between Oxidative Stress and Inflammatory Cytokines in Diabetic Nephropathy

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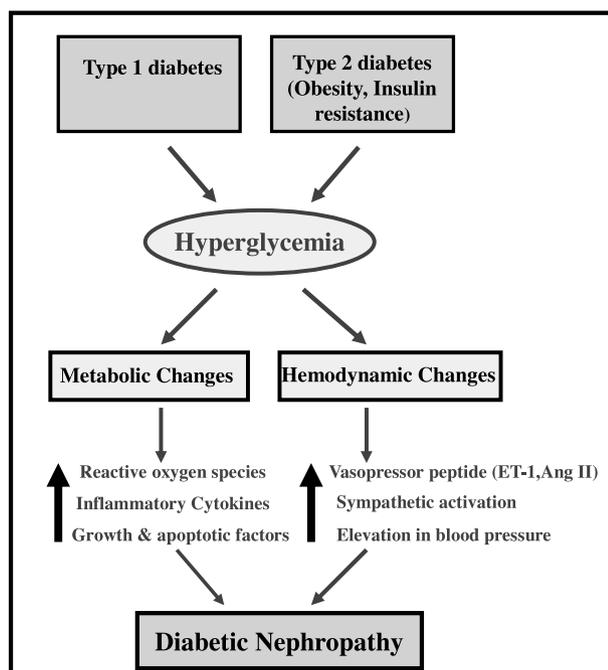
## SUMMARY

The prevalence of diabetes has dramatically increased worldwide due to the vast increase in the obesity rate. Diabetic nephropathy is one of the major complications of type 1 and type 2 diabetes and it is currently the leading cause of end-stage renal disease. Hyperglycemia is the driving force for the development of diabetic nephropathy. It is well known that hyperglycemia increases the production of free radicals resulting in oxidative stress. While increases in oxidative stress have been shown to contribute to the development and progression of diabetic nephropathy, the mechanisms by which this occurs are still being investigated. Historically, diabetes was not thought to be an immune disease; however, there is increasing evidence supporting a role for inflammation in type 1 and type 2 diabetes. Inflammatory cells, cytokines, and profibrotic growth factors including transforming growth factor- $\beta$  (TGF- $\beta$ ), monocyte chemoattractant protein-1 (MCP-1), connective tissue growth factor (CTGF), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin-1 (IL-1), interleukin-6 (IL-6), interleukin-18 (IL-18), and cell adhesion molecules (CAMs) have all been implicated in the pathogenesis of diabetic nephropathy via increased vascular inflammation and fibrosis. The stimulus for the increase in inflammation in diabetes is still under investigation; however, reactive oxygen species are a primary candidate. Thus, targeting oxidative stress-inflammatory cytokine signaling could improve therapeutic options for diabetic nephropathy. The current review will focus on understanding the relationship between oxidative stress and inflammatory cytokines in diabetic nephropathy to help elucidate the question of which comes first in the progression of diabetic nephropathy, oxidative stress, or inflammation.

## Introduction

Diabetic nephropathy is one of the most common microvascular complications of type 1 and type 2 diabetes mellitus and the leading cause of end-stage renal disease worldwide [1,2]. Many factors contribute to the development of diabetic nephropathy including hyperglycemia, hypertension, obesity, a sedentary lifestyle, hereditary, smoking, and advancing age [3,4]. Diabetic nephropathy is characterized by morphological and ultrastructural changes in the kidney including expansion of the molecular matrix and loss of the charge barrier on the glomerular basement membrane [5,6]. The progression from normal albuminuria to microalbuminuria is considered the initial step in diabetic nephropathy which further progresses to macroalbuminuria as renal function continues to deteriorate and glomerular filtration rate (GFR) starts to decline [5,6].

Careful control of glycemic status minimizes the symptoms of diabetic complications indicating that hyperglycemia is the main driving force behind the development of diabetic complications including diabetic nephropathy (Figure 1); however, strict glycemic control is difficult to maintain [7,8]. Current treatment strategies for diabetic nephropathy include glycemic and blood pressure control, low-protein diet, lipid-lowering drugs, and interference with the renin-angiotensin system [9,10]. Although these therapeutic options slow the progression of diabetic nephropathy, the burden and mortality rate of the disease remains very high and the majority of patients with diabetic nephropathy continue to progress to end-stage renal disease. Therefore, more detailed understanding of the molecular mechanisms for disease progression is needed. Both oxidative stress and inflammation are intimately linked with the development of diabetic nephropathy [1,10–12]. Increases in oxidative stress can increase the production of inflammatory



**Figure 1** Hyperglycemia is the main sign of type 1 and type 2 diabetes. In turn, hyperglycemia results in a myriad of metabolic and hemodynamic changes that are intimately associated with vascular complications of diabetes including diabetic nephropathy.

cytokines and likewise, an increase in inflammatory cytokines can stimulate the production of free radicals. The goal of this review is to examine how these two pathways interact to contribute to the development of diabetic nephropathy.

## Oxidative Stress in Diabetic Nephropathy

Under normal physiological conditions, there is a balance in the generation of oxygen-free radicals and the antioxidant defense mechanisms used to deactivate free radical toxicity [13–15]. Impairment in the oxidant/antioxidant equilibrium results in oxidative stress in numerous pathological conditions including diabetes leading to cellular damage [13–15]. Increasing evidence in both experimental and clinical studies suggests that there is a close link between hyperglycemia, oxidative stress, and diabetic complications [16,17]. Increased oxidative stress in diabetes likely contributes to the pathogenesis of diabetic nephropathy and its progression to end-stage renal disease [18–20]. Enhanced reactive oxygen species (ROS) production in experimental and clinical diabetes have been linked to vasoconstriction, vascular smooth muscle cell growth and migration, endothelial dysfunction, modification of extracellular matrix (ECM) proteins, and increased renal sodium reabsorption [21–24]. The importance of oxidative stress in diabetic nephropathy is underscored by the finding that inhibition of oxidative stress ameliorates the manifestations associated with streptozotocin-induced diabetic nephropathy [21,25]. Streptozotocin selectively targets and kills the beta cells of the pancreas resulting in an experimental model of type 1 diabetes mellitus. In

addition, overexpression of human cytoplasmic  $\text{Cu}^{2+}/\text{Zn}^{2+}$  superoxide dismutase (SOD-1) in streptozotocin-induced diabetic transgenic mice attenuates diabetic renal injury [26].

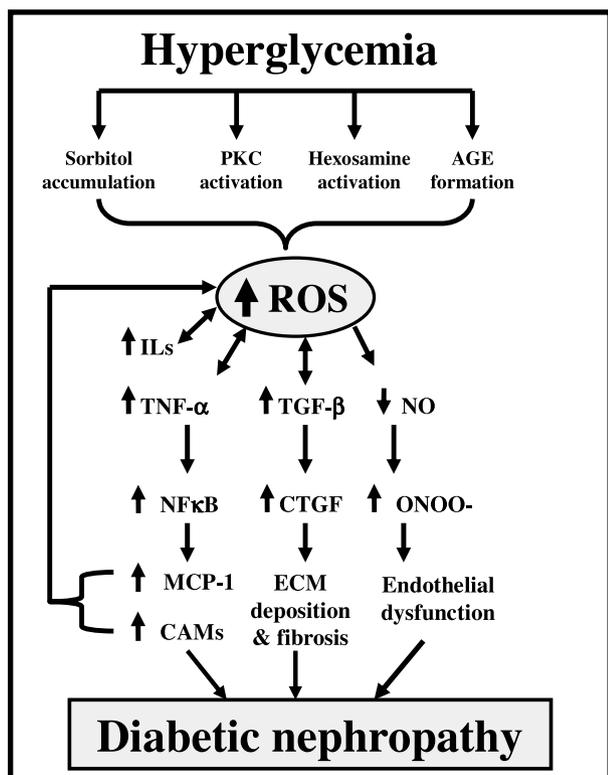
Renal damage and increased levels of oxidative stress are likely related to increased cellular glucose since hyperglycemia induces ROS generation. Reestablishing glycemic control soon after the induction of type 1 diabetes in rats and dogs prevents proteinuria, kidney hypertrophy, and the increase in oxidative and nitrate stress [27,28]. Careful glycemic control also ameliorates microalbuminuria in diabetic patients, and the risk of developing nephropathy is reduced [29]. Recent evidence suggests that nicotinamide adenine dinucleotide phosphate (NADPH) oxidase is the primary source of vascular and renal ROS production [30–36]; however, other possible sources include glucose-6-phosphate dehydrogenase, flux through the sorbitol/polyol pathway, and glycation of free amino groups on proteins and amino acids, mitochondrial electron transport enzymes, xanthine oxidase, cyclooxygenase, lipoxygenase, and uncoupled nitric oxide synthase [30,34,37]. For more information regarding the sources and role of oxidative stress in diabetes please see the following reviews [24,37–40].

## Inflammation in Diabetic Nephropathy

Diabetic nephropathy has traditionally been considered a non-immune disease; however, recent evidence shows an increase in macrophage infiltration and overproduction of leukocyte adhesion molecules in kidneys from diabetic humans and in experimental animal models of diabetes [41–45]. As a result, there is growing support for the notion that inflammation plays a key role in the pathogenesis of diabetic nephropathy. Leukocytes, monocytes, and macrophages have all been implicated in the process of diabetic nephropathy [41–45] and circulating inflammatory markers and proinflammatory cytokines are strongly associated with the risk of developing of diabetic complications [44–49]. Further support for inflammation to contribute to diabetes comes from studies where immunosuppressive strategies reduce renal macrophage accumulation and attenuate the development of diabetic nephropathy [50–52]. Although several recent reviews have examined the role of inflammatory cytokines in diabetic nephropathy [52–54], the focus of this review is on the interaction of oxidative stress and cytokines in diabetic nephropathy.

## Oxidative Stress Stimulates Cytokine Production

Oxidative stress can increase cytokine production via several different mechanisms. Oxygen derivatives, acting as second messengers, activate the transcription factors nuclear factor kappa B (NF $\kappa$ B) and activator protein-1 (AP-1) leading to the transcription of genes encoding cytokines, growth factors, and ECM proteins [55–58]. NF $\kappa$ B is suggested to play an important role in mesangial cell activation leading to renal injury and NF $\kappa$ B expression is increased in kidneys of diabetic experimental animals [59,60]. AP-1 mediates high glucose-induced TGF- $\beta$  production in mesangial cells and mutation of the AP-1 binding sites on TGF- $\beta$  abolished high glucose-mediated increases in TGF- $\beta$  levels [61].



**Figure 2** Schematic diagram showing the proposed relationship between hyperglycemia, oxidative stress, and inflammatory cytokines in the pathogenesis of diabetic nephropathy. ROS, reactive oxygen species; TNF- $\alpha$ , tumor necrosis factor- $\alpha$ ; NF $\kappa$ B, nuclear factor kappa B; MCP-1, monocyte chemoattractant protein-1; CAMs, cellular adhesion molecules; NO, nitric oxide; ONOO $^-$ , peroxynitrite; TGF- $\beta$ , transforming growth factor- $\beta$ ; CTGF, connective tissue growth factor; ILs, interleukins.

Peroxyntirite has also been implicated in the enhanced inflammation in diabetes as the result of decreased nitric oxide bioavailability [14,62]. In addition, enhanced macrophage migration in diabetes induces the release of inflammatory and profibrotic cytokines, which further stimulates ROS production [11,63]. Therefore, oxidative stress-induced cytokine production is likely to further increase oxidative stress levels setting up a vicious cycle (Figure 2). The remainder of this review focuses on the relationship between oxidative stress and the most commonly involved cytokines in the progression of diabetic nephropathy.

### Transforming Growth Factor- $\beta$ (TGF- $\beta$ )

TGF- $\beta$  is a hypertrophic and fibrogenic cytokine and the causative agent of mesangial expansion and renal insufficiency in human and type 1 experimental models of diabetic nephropathy [64–66]. TGF- $\beta$  stimulates the deposition of ECM via direct upregulation of matrix protein genes, inhibition of matrix degradation by suppressing proteases, and increasing the synthesis of protease inhibitors, increasing cell surface expression of integrins to promote attachment to newly synthesized matrix, and auto-inducing its

own production [67,68]. TGF- $\beta$ 1 mRNA and protein levels are increased in both glomerular and tubular compartments of various rat and mouse experimental models of type 1 and type 2 diabetes [66,69–71]. Direct support for the key role played by TGF- $\beta$  in the development of diabetic nephropathy comes from studies where treatment with neutralizing monoclonal antibodies to TGF- $\beta$  prevents glomerular hypertrophy, mesangial matrix expansion, and glomerulosclerosis and preserves renal function in streptozotocin-induced type 1 diabetes and “db/db” type 2 diabetic mice [72,73]. While the role of TGF- $\beta$  in diabetes is relatively well established, there remains some question as to the stimulus to increase TGF- $\beta$  levels in the diabetic kidney. We propose that the increase is secondary to a glucose-induced increase in ROS production.

Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) increases TGF- $\beta$  protein synthesis and stimulates collagen and fibronectin gene expression in cultured human mesangial cells [74]. H<sub>2</sub>O<sub>2</sub> also increases TGF- $\beta$  mRNA and collagen expression in cultured NRK-49F fibroblasts and rat mesangial cells [58]. H<sub>2</sub>O<sub>2</sub> effects on ECM protein expression in mesangial cells are directly mediated by TGF- $\beta$  as incubation of mesangial cells with an anti-TGF- $\beta$  antibody blocked the effects of H<sub>2</sub>O<sub>2</sub> on ECM gene expression [74]. However, H<sub>2</sub>O<sub>2</sub> has also been shown to activate NF $\kappa$ B in Jurkat T cells [56]. Therefore, it is possible that the effect of H<sub>2</sub>O<sub>2</sub> on TGF- $\beta$  is mediated by NF $\kappa$ B activation. H<sub>2</sub>O<sub>2</sub> is unlikely to be the sole ROS that stimulates TGF- $\beta$  levels. High glucose-induced increases in TGF- $\beta$  and fibronectin gene expression in rat mesangial cells is blocked by incubation with SOD and an NADPH oxidase selective inhibitor but not catalase, suggesting that in rat cells superoxide and not H<sub>2</sub>O<sub>2</sub> stimulates TGF- $\beta$  [75]. Incubation of rat glomerular mesangial cells with advanced oxidation protein products (AOPP) increases superoxide, TGF- $\beta$ , and ECM protein levels, and this effect is blocked by SOD [76]. AOPPs are cross-linking protein products formed during oxidative stress and AOPP levels are increased in plasma and renal homogenates of streptozotocin-induced type 1 diabetic male Sprague-Dawley rats [35]. In diabetic streptozotocin rats, treatment with AOPP further increases oxidative stress and TGF- $\beta$  levels, both of which were blocked by NADPH oxidase inhibition [35]. Therefore, while evidence supports the hypothesis that increased inflammation in the diabetic kidney is secondary to an increase in oxidative stress, whether this is a direct or an indirect effect remains to be elucidated.

Although data supporting a direct temporal relationship between increased ROS and TGF- $\beta$  stimulation of ECM overproduction in diabetes *in vivo* is scarce, there is indirect evidence. Antioxidant treatment decreases TGF- $\beta$  levels in the diabetic kidney. Mice overexpressing SOD1 are protected from streptozotocin-induced increases in renal TGF- $\beta$  and ECM production compared to diabetic wild-type mice [26] and treatment of streptozotocin diabetic male Wistar rats with SOD-PEG decreases renal 8-hydroxyl guanosine (8-OHdG), a marker of oxidative stress, TGF- $\beta$ , and fibronectin protein levels [75]. Similarly, treatment with an SOD mimetic inhibits glomerular matrix expansion via the suppression of TGF- $\beta$  in female Sprague-Dawley rats following streptozotocin injection [76]. The potential importance of SOD in regulating oxidative stress and TGF- $\beta$  levels in the diabetic kidney were underscored by studies on SOD1 knockout mice, where streptozotocin resulted in greater increases in renal TGF- $\beta$  in

knockout mice compared to wild-type littermates and this increase was blocked by a SOD mimetic [77]. Thioredoxin has also been implicated as an important determinant of redox balance in diabetes. The thioredoxin system reduces ROS through reversible oxidation of thioredoxin. Transgenic mice over-expressing thioredoxin-1 have less renal damage, lower 8-OHdG excretion, and suppression of TGF- $\beta$  mRNA following streptozotocin treatment compared to wild-type streptozotocin mice independent of glycemic status [78]. Additional evidence suggests that thioredoxin activity may be suppressed in diabetes. High glucose increases thioredoxin interacting protein (Txnip), an inhibitor of thioredoxin, in proximal tubule HK-2 cells, human aortic smooth muscle cells, and in male Ren-2 and Sprague-Dawley rats following streptozotocin injection [79,80]. In cultured human aortic smooth muscle cells, adenoviral overexpression of thioredoxin reduces glucose-induced ROS while overexpression of Txnip increases ROS. In addition, gene knockdown of Txnip reduces ROS in streptozotocin diabetic male Sprague-Dawley rats [80]. Finally, both vitamin E and taurine supplementation decrease oxidative stress levels and glomerular TGF- $\beta$  expression in streptozotocin-induced diabetic male Sprague-Dawley rats [81,82]. These data all support the hypothesis that oxidative stress significantly contributes to the increase in TGF- $\beta$  in diabetic nephropathy (Figure 2).

### Connective Tissue Growth Factor

Connective tissue growth factor (CTGF) is a highly profibrogenic molecule which is overexpressed in many fibrotic lesions [83,84]. CTGF is transcriptionally activated by TGF- $\beta$  and is considered the major downstream effector of TGF- $\beta$  [83,84]. CTGF is the key factor in stimulating connective tissue cell proliferation, ECM production, and other profibrotic properties of TGF- $\beta$  [85,86]. CTGF and TGF- $\beta$  exhibit shared fibrogenic and angiogenic properties *in vivo* as they both promote cell adhesion, migration, proliferation, and differentiation [85,86]. Thus, CTGF plays an important role in collagen production and the maintenance of fibrotic lesions.

Recent studies suggest that CTGF is critically involved in the pathogenesis and progression of diabetic nephropathy [84]. High glucose concentrations and advanced glycation end products stimulate CTGF and TGF- $\beta$  production in cultured mesangial cells and both are involved in diabetes-induced increases in ECM production, cell migration, and fibrosis [86–89]. CTGF is upregulated in experimental models of type 1 diabetic nephropathy and in diabetic patients [90–93]. In streptozotocin-induced diabetes, urinary CTGF increases during the early development of clinical symptoms then decreases as animals become proteinuric [94]. Urinary CTGF is also elevated in diabetic patients with albuminuria, which is prognostic for the progression to microalbuminuria [95]. Furthermore, inhibition of CTGF signaling using antisense preserves the structure and function of kidney in mouse models of type 1 and type 2 diabetes, underscoring the importance of CTGF in the pathogenesis of diabetic nephropathy [96].

The exact relationship between CTGF and oxidative stress remains to be investigated; however, advanced glycation end products induce oxidative stress and CTGF production [97–99]. In many experimental models of type 1 and type 2 diabetic nephropathy, elevations in oxidative stress parallel elevations in

CTGF levels [100–102]. Enhanced renal NADPH oxidase activity in streptozotocin-induced type 1 diabetic mice is associated with increased renal TGF- $\beta$ , CTGF, and collagen IV expression [102]. However, NADPH oxidase activity and expression are unchanged in diabetic protein kinase C (PKC)-beta knock out mice despite a decrease in renal TGF- $\beta$ , CTGF, and collagen IV expression [102]. We postulate that increased oxidative stress during diabetes is the trigger for stimulation of TGF- $\beta$ -CTGF fibrotic signaling (Figure 2).

### Interleukins (ILs)

#### IL-1

IL-1 increases the expression of chemotactic factors and adhesion molecules, enhances vascular endothelial permeability, and stimulates the proliferation of mesangial cells and matrix synthesis [53,54]. IL-1 was first implicated in the development of diabetic nephropathy when glomerular basement proteins isolated from streptozotocin-induced diabetic male rats had significantly greater macrophage, TNF- $\alpha$ , and IL-1 production compared to control rats [103]. IL-1 levels are also elevated in kidneys from male Sprague-Dawley rats treated with streptozotocin compared to control rats [104] and renal expression of IL-1 is significantly correlated with urinary albumin excretion [105].

#### IL-6

Renal IL-6 expression is positively related to mesangial proliferation, tubular atrophy in diverse models of renal disease, supporting the role of IL-6 in the progression of renal disease [54]. Serum IL-6 levels are significantly higher in patients with type 2 diabetic nephropathy compared to levels observed in diabetic patients without nephropathy suggesting a role for IL-6 in the pathogenesis of diabetic nephropathy [106–108]. In fact, serum IL-6 levels are similar in type 2 diabetic patients with normal albumin excretion and microalbuminuria, but significantly increased in patients with diabetic nephropathy and clinical albuminuria [109]. Serum IL-6 levels are also significantly greater in diabetic patients with overt proteinuria compared to normoalbuminuric and microalbuminuric patients [110,111]. Using high-resolution *in situ* hybridization in kidney biopsies of Japanese patients with diabetic nephropathy, it was found that interstitial expression of IL-6 mRNA correlated significantly with the degree of interstitial injury [112]. Clinical reports are in agreement with studies in experimental animals where renal expression of IL-6 mRNA is increased in streptozotocin diabetic rats compared to controls and levels are significantly associated with urinary albumin excretion [105]. These data support a role for IL-6 in the progression of diabetic nephropathy in the later stages of the disease.

#### IL-18

IL-18 is a potent inflammatory cytokine secreted from activated monocytes/macrophages and it is known to induce interferon- $\gamma$  (IFN- $\gamma$ ), which increases functional chemokine receptor expression in human mesangial cells [53]. IL-18 also stimulates the production of other inflammatory cytokines including IL-1, TNF- $\alpha$ ,

and IL-6, upregulates ICAM-1 expression, and induces endothelial cell apoptosis [53,108]. This raises the possibility that an increase in IL-18 in diabetic nephropathy may precede the observed increase in IL-6. IL-18 is constitutively expressed in renal tubular epithelia, and infiltrating monocytes, macrophages and proximal tubular cells have all been identified as potential sources of IL-18 production [53,113]. Expression of IL-18 is increased in renal biopsies from patients with diabetic nephropathy in proximal and epithelial tubular cells [114] and patients with type 2 diabetes have significantly higher serum and urinary levels of IL-18 compared to healthy controls [108,109,115,116]. Moreover, there is a positive correlation between IL-18 levels in diabetic patients and the development of urinary albumin excretion, with the highest IL-18 levels found in patients with microalbuminuria and clinical albuminuria [108,109,116,117].

### **Oxidative Stress Stimulates IL Production**

Despite increasing evidence supporting a role for ILs in diabetic nephropathy, little is known regarding the stimulus for increased IL production; however, there is evidence to support a role for oxidative stress. Hyperglycemia-induced oxidative stress and advanced glycation end products have been suggested to induce inflammatory cytokines. In healthy volunteers with either normal or impaired glucose tolerance, acute hyperglycemia increases plasma IL-6 and IL-18 concentrations which are blocked by infusion of the antioxidant glutathione 5 min prior to the glucose infusion, indicating that oxidative stress mediates the increase in IL production [111]. In addition, in older diabetic patients with poor glycemic control and asymptomatic individuals with abnormal fasting glycemia, thiobarbituric acid-reactive substances a marker of oxidative stress, and IL-6 levels are independently correlated with C-reactive protein, suggesting that oxidative stress promotes a state of low-grade systemic inflammation in elderly patients with type II diabetes [118,119]. In hyperglycemia, increased oxidative stress results in an increase in the formation and deposition of advanced glycation end products and their receptors (RAGE) in tissues and RAGE stimulation induces the activation of NF $\kappa$ B which may lead to an increase in IL production [120,121]. Also, as noted above, oxidative stress can directly activate the transcription factors NF $\kappa$ B and AP-1 [55,56] leading to the transcription of ILs either directly or through the induction of other cytokines. In support of this scheme, increased IL-1 production from macrophages incubated with glomerular basement proteins isolated from streptozotocin-induced diabetic male rats was found to be advanced glycation end products-dependent [103].

There is also evidence suggesting that IL production is increased in response to increased oxidative stress via the stimulation of alternative pathways in diabetes. In diabetic nephropathy patients, angiotensin receptor blockers suppress oxidative stress and inflammation and provide protection against the progression of diabetic nephropathy [122]. In agreement with this finding, treatment of streptozotocin-induced diabetic male Sprague-Dawley rats with the ACE inhibitor enalapril prevents enhanced IL-6 expression, leading to a decrease in urinary cytokine excretion and a reduction in albuminuria [105]. Angiotensin converting enzyme inhibitors

have been shown to decrease levels of oxidative stress in various experimental models. Alternatively, treatment of human renal proximal tubular epithelial cells with TGF- $\beta$  increases IL-18 mRNA expression [114], and TGF- $\beta$  is known to be increased by oxidative stress. These data suggest that elevated oxidative stress during diabetes is a potential mediator for increased ILs production (Figure 2).

### **Tumor Necrosis Factor-alpha (TNF- $\alpha$ )**

TNF- $\alpha$  is a pleiotropic cytokine produced mainly in macrophages and monocytes and is involved in systemic inflammation [123,124]. TNF- $\alpha$  induces a local inflammatory response by initiating a cascade of cytokines and increasing vascular permeability, thereby recruiting macrophage and neutrophils to a site of infection [123,124]. TNF- $\alpha$  activates NF $\kappa$ B signaling mediating the transcription of various cytokines involved in cell survival and proliferation, inflammatory responses and cell adhesion, and antiapoptotic factors [125–129]. Because TNF- $\alpha$  is cytotoxic to glomerular, mesangial, and epithelial cells and can induce renal damage [130], it has been shown to play a pathophysiological role in several experimental models of renal disease including lupus nephritis, crescentic glomerulonephritis, mesangial proliferative glomerulonephritis, diabetes, hypertension, and the remnant kidney model of nephropathy [130–133].

A role for TNF- $\alpha$  in diabetic nephropathy is supported by the finding that urinary albumin excretion significantly correlates with renal TNF- $\alpha$  levels and urinary TNF- $\alpha$  excretion in streptozotocin-induced diabetic rats [134,135]. Moreover, the increase in renal TNF- $\alpha$  levels and excretion precede the increase in albuminuria in diabetes. Urinary TNF- $\alpha$  levels are also elevated in type 2 diabetic patients and TNF- $\alpha$  levels rise as diabetic nephropathy progresses, suggesting that increased TNF- $\alpha$  levels contribute to the development of renal damage [135,136]. TNF- $\alpha$  also contributes to sodium retention and renal hypertrophy, which are early characteristic signs of streptozotocin-induced diabetic nephropathy [137]. Renal TNF- $\alpha$  expression, particularly in the glomerulus and tubulointerstitium, is increased in streptozotocin diabetic rat kidneys, and serum TNF- $\alpha$  is increased in type 2 diabetic patients [134,136]. Therefore, TNF- $\alpha$  plays an important role in the incidence and progression of diabetic nephropathy and renal TNF- $\alpha$  levels correlate with markers of diabetic nephropathy.

The relationship between oxidative stress and TNF- $\alpha$  is complex. TNF- $\alpha$  has been shown to increase ROS and ROS have been shown to increase TNF- $\alpha$  levels [138–140]. In the streptozotocin diabetic rat kidney, elevations in TNF- $\alpha$  increase oxidative stress leading to increased albumin permeability and urinary albumin excretion, a common marker of renal injury [140]. Additionally, elevated peroxynitrite levels are associated with increased TNF- $\alpha$  levels and increased glomerular lesion in streptozotocin diabetic rats [15]. These data suggest that TNF- $\alpha$  is upstream of oxidative stress in diabetic nephropathy. In contrast, administration of an SOD mimetic reduces renal TNF- $\alpha$  levels and albuminuria in type 2 diabetic Zucker rats [141], and the antioxidant tocotrienol offers reno-protection to streptozotocin diabetic rats via decreasing oxidative stress and modulating TNF- $\alpha$  and TGF- $\beta$ -induced inflammation [140]. These studies suggest that oxidative stress is

upstream of TNF- $\alpha$  activation in diabetic nephropathy (Figure 2). Thus, it is difficult to separate oxidative stress and TNF- $\alpha$  in diabetes. Further experimental and clinical studies using antioxidants and/or TNF- $\alpha$  inhibitors are required to determine the relationship between oxidative stress and TNF- $\alpha$  in diabetic nephropathy.

### Monocyte Chemoattractant Protein-1 (MCP-1)

Increasing evidences suggest that recruitment of inflammatory cells from the circulation into renal tissue plays a pivotal role in the progression of diabetic nephropathy. In particular, infiltration of activated T cells and monocytes initiate renal damage and eventually lead to a progressive loss of renal function. Chemokine (C-C motif) ligand 2 (CCL2) is a small cytokine belonging to the CC chemotactic chemokine family that is also known as monocyte chemoattractant protein-1 (MCP-1) [142–144]. MCP-1 recruits monocytes, memory T cells, and dendritic cells to sites of tissue injury and infection. MCP-1 is expressed by monocytes, vascular endothelial cells, smooth muscle cells, glomerular mesangial cell, and osteoblastic cells. Many cytokines have been shown to stimulate the production of MCP-1 including IL-1, TNF- $\alpha$ , and TGF- $\beta$  [142–145].

Studies suggest that MCP-1 plays a role in the progression of diabetic renal injury. Cultured mesangial cells, podocytes, and renal tubular epithelial cells produce MCP-1 in the presence of high glucose and advanced glycation end products [146–149]. In streptozotocin-induced diabetes, MCP-1 is upregulated in glomeruli and tubulointerstitium and this increase contributes to renal fibrosis [150,151]. MCP-1-mediated macrophage infiltration contributes to the progression of diabetic nephropathy [95,146], as evidenced by the ability of MCP-1 inhibition to ameliorate the development of diabetic nephropathy in diabetic mice [146,152]. Clinically, urinary MCP-1 excretion and MCP-1 levels in renal biopsies are elevated in diabetic patients [153,154]. Furthermore, elevated renal MCP-1 in diabetic patients is associated with macrophage recruitment, albuminuria, tubulointerstitial injury, and the progression of diabetic nephropathy [152–155]. Elevated MCP-1 excretion in diabetic patients correlates with macroalbuminuria and was prognostic for deterioration of kidney function [93].

The relationship between oxidative stress and MCP-1 in diabetic nephropathy is unclear and requires further elucidation. However, in experimental animals, elevations in oxidative stress increase macrophage recruitment and renal ICAM-1 and MCP-1 expression in type 1 diabetic rats [156]. In addition, mesangial cells from streptozotocin-induced diabetic mice exhibit an increase in oxidative stress and inflammatory cytokines including MCP-1, and antioxidant treatment reduces MCP-1 levels [17]. Clinically, oxidative stress and plasma MCP-1 are significantly elevated in type 1 diabetic patients with microalbuminuria and poor glycemic control when compared with healthy control subjects [157] and increased oxidative stress is associated with elevations in MCP-1 expression in circulating monocytes in type 1 diabetic patients [158]. Plasma MCP-1 is also positively correlated with plasma malondialdehyde (MDA), a marker of oxidative stress, and albuminuria and vitamin E treatment reduced plasma MCP-1 and albuminuria in type

1 diabetic patients [157,158]. Collectively, these data suggest that oxidative stress-induced MCP-1 expression may contribute to the progression of diabetic nephropathy (Figure 2).

### Cell Adhesion Molecules (CAMs)

Vascular cell adhesion molecule-1 (VCAM-1), also known as CD106, is expressed on both large and small vessels following endothelial cell stimulation by cytokines such as TNF- $\alpha$  and IL-1 [159–161]. VCAM-1 promotes the adhesion of lymphocytes, monocytes, eosinophils, and basophils to the vascular endothelium. VCAM-1 also functions in leukocyte-endothelial cell signal transduction, and has been implicated in the development of cardiovascular diseases including diabetes [159–161]. Endothelial VCAM-1 expression increased in spontaneously diabetic KKAy mice [161]. Clinically, circulating VCAM-1 levels are elevated in patients with diabetic nephropathy [162] and diabetic patients with albuminuria have high plasma VCAM-1 levels and an increased risk of end-stage renal disease and death [163].

Inter-cellular adhesion molecule-1 (ICAM-1), also known as CD54, is a member of the immunoglobulin superfamily, which includes antibodies and T-cell receptors [164–167]. ICAM-1 is a ligand for  $\beta_2$ -integrins, a receptor found on leukocytes. When activated, ICAM-1 binds  $\beta_2$ -integrins on leukocyte cell surfaces promoting leukocyte adhesion to the endothelium and transmigration [164–167]. Like VCAM-1, ICAM-1 can be expressed by vascular endothelium, macrophages, and lymphocytes and can also be induced by cytokines such as IL-1 and TNF- $\alpha$  [164–168]. NF $\kappa$ B also increases gene transcription of ICAM-1 [164–166].

Hyperglycemia upregulates endothelial ICAM-1 expression [169] and a direct association between renal ICAM-1 expression and progressive renal injury has been shown in experimental animal models of renal diseases including diabetic nephropathy [166–171]. In streptozotocin-induced diabetic rats, anti-ICAM-1 antibody blocks renal macrophage infiltration suggesting that ICAM-1 mediates macrophage infiltration into the diabetic kidney [172]. Furthermore, ICAM-1 deficient “db/db” mice (ICAM-1<sup>-/-</sup>) have a decrease in glomerular macrophage infiltration and are protected from renal injury when compared to ICAM-1<sup>+/+</sup> “db/db” mice [173], confirming the notion that ICAM-1 is critically involved in diabetic nephropathy. Clinically, macrophage infiltration and ICAM-1 expression are elevated in kidneys of patients with diabetic nephropathy [166] and patients with type 1 and type 2 diabetes have elevated plasma and urinary ICAM-1 concentrations compared to subjects without renal injury [174–176].

There is evidence to support the hypothesis that CAM expression is stimulated by oxidative stress. It is well known that oxidative stress stimulates NF $\kappa$ B-induced CAMs expression. Advanced glycation end product-RAGE interactions also increase ROS formation in mesangial and endothelial cells with subsequent activation of NF $\kappa$ B and the release of inflammatory cytokines including ICAM-1 and VCAM-1 [159]. Diabetic “db/db” mice have elevated albuminuria, oxidative stress, and tubulointerstitial injury together with increased tubuloepithelial ICAM-1 expression [177]. Streptozotocin-induced diabetic rats have higher renal NADPH oxidase expression and urinary lipid peroxidation product

(LPO) in conjunction with increased glomerular ICAM-1 expression [178], and antioxidant treatment with taurine or NADPH oxidase inhibition reduces renal damage and ICAM-1 expression [178,179]. Immunosuppressive treatment with mycophenolate mofetil (MMF) also ameliorates early renal injury via the inhibition of oxidative stress-induced ICAM-1, MCP-1, and TGF- $\beta$  expression in streptozotocin-induced diabetic rat kidneys [17,156]. These data suggest that oxidative stress is upstream of ICAM-1 activation in the progression of diabetic nephropathy. However, a recent study demonstrated that xanthine oxidase inhibition reduced albuminuria and renal injury in diabetic “db/db” mouse kidneys without a reduction in renal oxidative stress [177]. Instead a reduction in tubular ICAM-1 expression and subsequent reduction in macrophage infiltration and inflammatory cytokines was identified as the mechanisms of reno-protective [177]. It is possible that there were localized decreases in oxidative stress in certain cell types or in other tissue that were not detected in the study.

## Perspectives

There is a close association between oxidative stress and inflammation in diabetes and we hypothesize that an increase in oxidative stress-derived inflammation is a major mechanism in the pathogenesis and progression of diabetic nephropathy. In addition, an increase in inflammatory cytokine levels in diabetes may drive a further increase in oxidative stress as renal injury becomes more pronounced setting up a vicious cycle. However, due to the complex and intimate association between increased oxidative stress and increased inflammation, dissecting the temporal nature of the relationship is a very difficult task. We are in the process of establishing an animal model of diabetic nephropathy that better reflects the human condition in which hyperglycemia, hypertension, proteinuria, and a decline in renal function are all present. We are currently studying streptozotocin-induced diabetic spontaneously hypertensive rats (SHR). These rats are hypertensive, hyperglycemic, and albuminuric with reduced renal function. We will use this model of type 1 diabetes to better define what comes first in diabetic nephropathy, oxidative stress or inflammation. A better understanding of the relationship between oxidative stress and inflammatory cytokines in the progression of diabetic nephropathy will facilitate the development of new treatment options and improve current therapeutic strategies.

## Conflict of Interest

The authors have no conflict of interest.

## References

1. Lopes AA. End-stage renal disease due to diabetes in racial/ethnic minorities and disadvantaged populations. *Ethn Dis* 2009;19:S1(47–51). Review.
2. Ohga S, Shikata K, Yozai K, et al. Thiazolidinedione ameliorates renal injury in experimental diabetic rats through anti-inflammatory effects mediated by inhibition of NF-kappaB activation. *Am J Physiol Renal Physiol* 2007;292:F1141–F1150.
3. Rossing P. Diabetic nephropathy: Worldwide epidemic and effects of current treatment on natural history. *Curr Diab Rep* 2006;6:479–483.

4. Romero-Aroca P, Mendez-Marin I, Baget-Bernaldiz M, Fernández-Ballart J, Santos-Blanco E. Review of the relationship between renal and retinal microangiopathy in diabetes mellitus patients. *Curr Diabetes Rev* 2010; 6: 88–101. Review.
5. Hovind P, Rossing P, Tarnow L, Smidt UM, Parving HH. Progression of diabetic nephropathy. *Kidney Int* 2001;59:702–709.
6. Parving HH. Diabetic nephropathy: Prevention and treatment. *Kidney Int* 2001;60:2041–2055.
7. Kilpatrick ES, Rigby AS, Atkin SL. The diabetes control and complications trial: The gift that keeps giving. *Nat Rev Endocrinol* 2009;5:537–545. Review.
8. Lewko B, Stepinski J. Hyperglycemia and mechanical stress: Targeting the renal podocyte. *J Cell Physiol* 2009;221:288–295.
9. Battle D. Clinical and cellular markers of diabetic nephropathy. *Kidney Int* 2003;63:2319–2330.
10. Shah IM, Mackay SP, McKay GA. Therapeutic strategies in the treatment of diabetic nephropathy: A translational medicine approach. *Curr Med Chem* 2009;16:997–1016. Review.
11. Ziyadeh FN, Wolf G. Pathogenesis of the podocytopathy and proteinuria in diabetic glomerulopathy. *Curr Diabetes Rev* 2008;4:39–45. Review.
12. Jeong KH, Lee TW, Ihm CG, Lee SH, Moon JY, Lim SJ. Effects of sildenafil on oxidative and inflammatory injuries of the kidney in streptozotocin-induced diabetic rats. *Am J Nephrol* 2009;29:274–282.
13. Anzalone R, La Rocca G, Di Stefano A, et al. Role of endothelial cell stress in the pathogenesis of chronic heart failure. *Front Biosci* 2009;14:2238–2247. Review.
14. Zheng L, Kern TS. Role of nitric oxide, superoxide, peroxynitrite and PARP in diabetic retinopathy. *Front Biosci* 2009;14:3974–3987.
15. Xiao H, Li Y, Qi J, Wang H, Liu K. Peroxynitrite plays a key role in glomerular lesions in diabetic rats. *J Nephrol* 2009;22:800–808.
16. Brown WV. Microvascular complications of diabetes mellitus: Renal protection accompanies cardiovascular protection. *Am J Cardiol* 2008;102:10L–13L. Review.
17. Wu J, Mei C, Vlassara H, Striker GE, Zheng F. Oxidative stress-induced JNK activation contributes to proinflammatory phenotype of aging diabetic mesangial cells. *Am J Physiol Renal Physiol* 2009;297:F1622–F1631.
18. Szabo C. Role of nitrosative stress in the pathogenesis of diabetic vascular dysfunction. *Br J Pharmacol* 2009;156:713–727.
19. Wolf G. New insights into the pathophysiology of diabetic nephropathy: From haemodynamics to molecular pathology. *Eur J Clin Invest* 2004;34:785–796.
20. Kanwar YS, Wada J, Sun L, et al. Diabetic nephropathy: Mechanisms of renal disease progression. *Exp Biol Med* 2008;233:4–11.
21. Asaba K, Tojo A, Onozato ML, et al. Effects of NADPH oxidase inhibitor in diabetic nephropathy. *Kidney Int* 2005;67:1890–1898.
22. Ishii N, Patel KP, Lane PH, et al. Nitric oxide synthesis and oxidative stress in the renal cortex of rats with diabetes mellitus. *J Am Soc Nephrol* 2001;12:1630–1639.
23. Son SM, Whalin MK, Harrison DG, Taylor WR, Griendling KK. Oxidative stress and diabetic vascular complications. *Curr Diab Rep* 2004;4:247–252.
24. Satoh M, Fujimoto S, Haruna Y, et al. NAD(P)H oxidase and uncoupled nitric oxide synthase are major sources of glomerular superoxide in rats with experimental diabetic nephropathy. *Am J Physiol Renal Physiol* 2005;288:F1144–F1152.
25. Thallas-Bonke V, Thorpe SR, Coughlan MT, et al. Inhibition of NADPH oxidase prevents advanced glycation end product-mediated damage in diabetic nephropathy through a protein kinase C-alpha-dependent pathway. *Diabetes* 2008;57:460–469.
26. Craven PA, Melhem MF, Phillips SL, DeRubertis FR. Overexpression of Cu2+/Zn2+ superoxide dismutase protects against early diabetic glomerular injury in transgenic mice. *Diabetes* 2001;50:2114–2125.
27. Kern TS, Engerman RL. Arrest of glomerulopathy in diabetic dogs by improved glycaemic control. *Diabetologia* 1990;33:522–525.
28. Kowluru RA. Effect of reinstatement of good glycemic control on retinal oxidative stress and nitrate stress in diabetic rats. *Diabetes* 2003;52:818–823.
29. Bojestig M, Arnqvist HJ, Karlberg BE, Ludvigsson J. Glycemic control and prognosis in type I diabetic patients with microalbuminuria. *Diabetes Care* 1996;19:313–317.

30. Forbes JM, Coughlan MT, Cooper ME. Oxidative stress as a major culprit in kidney disease in diabetes. *Diabetes* 2008;**57**:1446–1454.
31. Tojo A, Asaba K, Onozato ML. Suppressing renal NADPH oxidase to treat diabetic nephropathy. *Expert Opin Ther Targets* 2007;**11**:1011–1018.
32. Gao L, Mann GE. Vascular NAD(P)H oxidase activation in diabetes: A double-edged sword in redox signalling. *Cardiovasc Res* 2009;**82**:9–20.
33. Inoguchi T, Li P, Umeda F, et al. High glucose level and free fatty acid stimulate reactive oxygen species production through protein kinase C-dependent activation of NAD(P)H oxidase in cultured vascular cells. *Diabetes* 2000;**49**:1939–1945.
34. Lee HB, Yu MR, Yang Y, Jiang Z, Ha H. Reactive oxygen species-regulated signaling pathways in diabetic nephropathy. *J Am Soc Nephrol* 2003;**14**:S241–S245.
35. Shi XY, Hou FF, Niu HX, et al. Advanced oxidation protein products promote inflammation in diabetic kidney through activation of renal nicotinamide adenine dinucleotide phosphate oxidase. *Endocrinology* 2008;**149**:1829–1839.
36. Gorin Y, Block K, Hernandez J, et al. Nox4 NAD(P)H oxidase mediates hypertrophy and fibronectin expression in the diabetic kidney. *J Biol Chem* 2005;**280**:39616–39626.
37. Elmarakby AA, Williams JM, Pollock DM. Targeting sources of superoxide and increasing nitric oxide bioavailability in hypertension. *Curr Opin Investig Drugs* 2003;**4**:282–290. Review.
38. Wei W, Liu Q, Tan Y, Liu L, Li X, Cai L. Oxidative stress, diabetes, and diabetic complications. *Hemoglobin* 2009;**33**:370–377. Review.
39. Pérez-Matute P, Zulet MA, Martínez JA. Reactive species and diabetes: Counteracting oxidative stress to improve health. *Curr Opin Pharmacol* 2009;**9**:771–779. Review.
40. Friederich M, Hansell P, Palm F. Diabetes, oxidative stress, nitric oxide and mitochondria function. *Curr Diabetes Rev* 2009;**5**:120–144. Review.
41. Chow F, Ozols E, Nikolic-Paterson DJ, Atkins RC, Tesch GH. Macrophages in mouse type 2 diabetic nephropathy: Correlation with diabetic state and progressive renal injury. *Kidney Int* 2004;**65**:116–128.
42. Chow FY, Nikolic-Paterson DJ, Atkins RC, Tesch GH. Macrophages in streptozotocin-induced diabetic nephropathy: Potential role in renal fibrosis. *Nephrol Dial Transplant* 2004;**19**:2987–2996.
43. Galkina E, Ley K. Leukocyte recruitment and vascular injury in diabetic nephropathy. *J Am Soc Nephrol* 2006;**17**:368–377.
44. Shikata K, Makino H. Role of macrophages in the pathogenesis of diabetic nephropathy. *Contrib Nephrol* 2001;**134**:46–54.
45. Nguyen D, Ping F, Mu W, Hill P, Atkins RC, Chadban SJ. Macrophage accumulation in human progressive diabetic nephropathy. *Nephrology* 2006;**11**:226–231.
46. King GL. The role of inflammatory cytokines in diabetes and its complications. *J Periodontol* 2008;**79**:1527–1534.
47. McCarty MF. Adjuvant strategies for prevention of glomerulosclerosis. *Med Hypotheses* 2006;**67**:1277–1296.
48. Catalán V, Gómez-Ambrosi J, Ramirez B, et al. Proinflammatory cytokines in obesity: Impact of type 2 diabetes mellitus and gastric bypass. *Obes Surg* 2007;**17**:1464–1474.
49. Kim CS, Park HS, Kawada T, et al. Circulating levels of MCP-1 and IL-8 are elevated in human obese subjects and associated with obesity-related parameters. *Int J Obes* 2006;**30**:1347–1355.
50. Wittmann S, Daniel C, Stief A, Vogelbacher R, Amann K, Hugo C. Long-term treatment of sirolimus but not cyclosporine ameliorates diabetic nephropathy in the rat. *Transplantation* 2009;**87**:1290–1299.
51. Rodriguez-Iturbe B, Quiroz Y, Shahkarami A, Li Z, Vaziri ND. Mycophenolate mofetil ameliorates nephropathy in the obese Zucker rat. *Kidney Int* 2005;**68**:1041–1047.
52. Wu Y, Dong J, Yuan L, et al. Nephron and podocin loss is prevented by mycophenolate mofetil in early experimental diabetic nephropathy. *Cytokine* 2008;**44**:85–91.
53. Navarro-Gonzalez JF, Mora-Fernandez C. The role of inflammatory cytokines in diabetic nephropathy. *J Am Soc Nephrol* 2008;**19**:433–442.
54. Rivero A, Mora C, Muros M, Garcia J, Herrera H, Navarro-Gonzalez JF. Pathogenic perspectives for the role of inflammation in diabetic nephropathy. *Clin Sci* 2009;**116**:479–492.
55. Meyer M, Schreck R, Baeuerle PA. H<sub>2</sub>O<sub>2</sub> and antioxidants have opposite effects on activation of NF-kappa B and AP-1 in intact cells: AP-1 as secondary antioxidant-responsive factor. *EMBO J* 1993;**12**:2005–2015.
56. Schreck R, Rieber P, Baeuerle PA. Reactive oxygen intermediates as apparently widely used messengers in the activation of the NF-kappa B transcription factor and HIV-1. *EMBO J* 1991;**10**:2247–2258.
57. Kim SJ, Angel P, Lafyatis R, et al. Autoinduction of transforming growth factor beta 1 is mediated by the AP-1 complex. *Mol Cell Biol* 1990;**10**:1492–1497.
58. Nath KA, Grande J, Croatt A, Haugen J, Kim Y, Rosenberg ME. Redox regulation of renal DNA synthesis, transforming growth factor-beta1 and collagen gene expression. *Kidney Int* 1998;**53**:367–381.
59. Massy ZA, Guijarro C, O'Donnell MP, et al. The central role of nuclear factor-kappa B in mesangial cell activation. *Kidney Int* 1999;**71**:S76–S79.
60. Guijarro C, Egido J. Transcription factor-kappa B (NF-kappa B) and renal disease. *Kidney Intl* 2001;**59**:415–424.
61. Weigert C, Sauer U, Brodbeck K, Pfeiffer A, Haring HU, Schleicher ED. AP-1 proteins mediate hyperglycemia-induced activation of the human TGF-beta1 promoter in mesangial cells. *J Am Soc Nephrol* 2000;**11**:2007–2016.
62. Liang JH, Li YN, Qi JS, Jia XX. Peroxynitrite-induced protein nitration is responsible for renal mitochondrial damage in diabetic rat. *J Endocrinol Invest* 2010;**33**:140–146.
63. Hohmeier HE, Tran VV, Chen G, Gasa R, Newgard CB. Inflammatory mechanisms in diabetes: Lessons from the beta-cell. *Int J Obes Relat Metab Disord* 2003;**27**(Suppl 3):S12–S16.
64. Reeves WB, Andreoli TE. Transforming growth factor beta contributes to progressive diabetic nephropathy. *Proc Natl Acad Sci U S A*; **97**:7667–7669.
65. Ziyadeh FN, Sharma K, Ericksen M, Wolf G. Stimulation of collagen gene expression and protein synthesis in murine mesangial cells by high glucose is mediated by autocrine activation of transforming growth factor-beta. *J Clin Invest* 1994;**93**:536–542.
66. Yamamoto T, Nakamura T, Noble NA, Ruoslahti E, Border WA. Expression of transforming growth factor beta is elevated in human and experimental diabetic nephropathy. *Proc Natl Acad Sci U S A* 1993;**90**:1814–1818.
67. Ziyadeh FN, Han DC. Involvement of transforming growth factor-beta and its receptors in the pathogenesis of diabetic nephropathy. *Kidney Int* 1997;**60**:S7–S11.
68. Border WA, Yamamoto T, Noble NA. Transforming growth factor beta in diabetic nephropathy. *Diabetes Metab Rev* 1996;**12**:309–339.
69. Sharma K, Ziyadeh FN. Renal hypertrophy is associated with upregulation of TGF-beta 1 gene expression in diabetic BB rat and NOD mouse. *Am J Physiol Renal Physiol* 1994;**267**:F1094–F1101.
70. Hill C, Flyvbjerg A, Gronbaek H, et al. The renal expression of transforming growth factor-beta isoforms and their receptors in acute and chronic experimental diabetes in rats. *Endocrinology* 2000;**141**:1196–1208.
71. Nakamura T, Fukui M, Ebihara I, et al. mRNA expression of growth factors in glomeruli from diabetic rats. *Diabetes* 1993;**42**:450–456.
72. Sharma K, Jin Y, Guo J, Ziyadeh FN. Neutralization of TGF-beta by anti-TGF-beta antibody attenuates kidney hypertrophy and the enhanced extracellular matrix gene expression in STZ-induced diabetic mice. *Diabetes* 1996;**45**:522–530.
73. Ziyadeh FN, Hoffman BB, Han DC, et al. Long-term prevention of renal insufficiency, excess matrix gene expression, and glomerular mesangial matrix expansion by treatment with monoclonal antitransforming growth factor-beta antibody in db/db diabetic mice. *Proc Natl Acad Sci U S A* 2000;**97**:8015–8020.
74. Iglesias-De La, Cruz MC, Ruiz-Torres P, et al. Hydrogen peroxide increases extracellular matrix mRNA through TGF-beta in human mesangial cells. *Kidney Int* 2001;**59**:87–95.
75. Lin CL, Wang FS, Kuo YR, et al. Ras modulation of superoxide activates ERK-dependent fibronectin expression in diabetes-induced renal injuries. *Kidney Int* 2006;**69**:1593–1600.
76. Asaba K, Tojo A, Onozato ML, Goto A, Fujita T. Double-edged action of SOD mimetic in diabetic nephropathy. *J Cardiovasc Pharmacol* 2007;**49**:13–19.
77. DeRubertis FR, Craven PA, Melhem MF. Acceleration of diabetic renal injury in the superoxide dismutase knockout mouse: Effects of tempol. *Metabolism* 2007;**56**:1256–1264.

78. Hamada Y, Miyata S, Nii-Kono T, et al. Overexpression of thioredoxin1 in transgenic mice suppresses development of diabetic nephropathy. *Nephrol Dial Transplant* 2007;**22**:1547–1557.
79. Qi W, Chen X, Gilbert RE, et al. High glucose-induced thioredoxin-interacting protein in renal proximal tubule cells is independent of transforming growth factor-beta1. *Am J Pathol* 2007;**171**:744–754.
80. Schulze PC, Yoshioka J, Takahashi T, He Z, King GL, Lee RT. Hyperglycemia promotes oxidative stress through inhibition of thioredoxin function by thioredoxin-interacting protein. *J Biol Chem* 2004;**279**:30369–30374.
81. Montero A, Munger KA, Khan RZ, et al. F(2)-isoprostanes mediate high glucose-induced TGF-beta synthesis and glomerular proteinuria in experimental type I diabetes. *Kidney Int* 2000;**58**:1963–1972.
82. Higo S, Miyata S, Jiang QY, Kitazawa R, Kitazawa S, Kasuga M. Taurine administration after appearance of proteinuria retards progression of diabetic nephropathy in rats. *Kobe J Med Sci* 2008;**54**:E35–E45.
83. Leask A, Abraham DJ. TGF-beta signaling and the fibrotic response. *FASEB J* 2004;**18**:816–827.
84. Nguyen TQ, Tarnow L, Jorsal A, et al. Plasma connective tissue growth factor is an independent predictor of end-stage renal disease and mortality in type I diabetic nephropathy. *Diabetes Care* 2008;**31**:1177–1182.
85. Perbal B. CCN proteins: Multifunctional signalling regulators. *Lancet* 2004;**363**:62–64.
86. Riser BL, Denichilo M, Cortes P, et al. Regulation of connective tissue growth factor activity in cultured rat mesangial cells and its expression in experimental diabetic glomerulosclerosis. *J Am Soc Nephrol* 2000;**11**:25–38.
87. Blom IE, van Dijk AJ, de Weger RA, Tilanus MG, Goldschmeding R. Identification of human ccn2 (connective tissue growth factor) promoter polymorphisms. *Mol Pathol* 2001;**54**:192–196.
88. Blom IE, van Dijk AJ, Wieten L, et al. In vitro evidence for differential involvement of CTGF, TGFbeta, and PDGF-BB in mesangial response to injury. *Nephrol Dial Transplant* 2001;**16**:1139–1148.
89. Burns WC, Twigg SM, Forbes JM, et al. Connective tissue growth factor plays an important role in advanced glycation end product-induced tubular epithelial-to-mesenchymal transition: Implications for diabetic renal disease. *J Am Soc Nephrol* 2006;**17**:2484–2494.
90. Adler SG, Kang SW, Feld S, et al. Glomerular mRNAs in human type I diabetes: Biochemical evidence for microalbuminuria as a manifestation of diabetic nephropathy. *Kidney Int* 2001;**60**:2330–2336.
91. Ito Y, Aten J, Bende RJ, et al. Expression of connective tissue growth factor in human renal fibrosis. *Kidney Int* 1998;**53**:853–861.
92. Roestenberg P, van Nieuwenhoven FA, Joles JA, et al. Temporal expression profile and distribution pattern indicate a role of connective tissue growth factor (CTGF/CCN-2) in diabetic nephropathy in mice. *Am J Physiol Renal Physiol* 2006;**290**:F1344–F1354.
93. Wahab NA, Schaefer L, Weston BS, et al. Glomerular expression of thrombospondin-1, transforming growth factor beta and connective tissue growth factor at different stages of diabetic nephropathy and their interdependent roles in mesangial response to diabetic stimuli. *Diabetologia* 2005;**48**:2650–2660.
94. Riser BL, Cortes P, DeNichilo M, et al. Urinary CCN2 (CTGF) as a possible predictor of diabetic nephropathy: Preliminary report. *Kidney Int* 2003;**64**:451–458.
95. Tam FW, Riser BL, Meeran K, Rambow J, Pusey CD, Frankel AH. Urinary monocyte chemoattractant protein-1 (MCP-1) and connective tissue growth factor (CCN2) as prognostic markers for progression of diabetic nephropathy. *Cytokine* 2009;**47**:37–42.
96. Guha M, Xu ZG, Tung D, Lanting L, Natarajan R. Specific down-regulation of connective tissue growth factor attenuates progression of nephropathy in mouse models of type 1 and type 2 diabetes. *FASEB J* 2007;**21**:3355–3368.
97. Forbes JM, Cooper ME, Oldfield MD, Thomas MC. Role of advanced glycation end products in diabetic nephropathy. *J Am Soc Nephrol* 2003;**14**(Suppl 3): S254–S258. Review.
98. Yan HD, Li XZ, Xie JM, Li M. Effects of advanced glycation end products on renal fibrosis and oxidative stress in cultured NRK-49F cells. *Chin Med J* 2007;**120**:787–793.
99. Forbes JM, Thallas V, Thomas MC, Founds HW, Burns WC, Jerums G, Cooper ME. The breakdown of preexisting advanced glycation end products is associated with reduced renal fibrosis in experimental diabetes. *FASEB J* 2003;**17**:1762–1764.
100. Lu Q, Yin XX, Wang JY, Gao YY, Pan YM. Effects of Ginkgo biloba on prevention of development of experimental diabetic nephropathy in rats. *Acta Pharmacol Sin* 2007;**28**:818–828.
101. Ohtomo S, Nangaku M, Izuhara Y, Takizawa S, Strihou CY, Miyata T. Cobalt ameliorates renal injury in an obese, hypertensive type 2 diabetes rat model. *Nephrol Dial Transplant* 2008;**23**:1166–1172.
102. Ohshiro Y, Ma RC, Yasuda Y, et al. Reduction of diabetes-induced oxidative stress, fibrotic cytokine expression, and renal dysfunction in protein kinase C beta-null mice. *Diabetes* 2006;**55**:3112–3120.
103. Hasegawa G, Nakano K, Sawada M, et al. Possible role of tumor necrosis factor and interleukin-1 in the development of diabetic nephropathy. *Kidney Int* 1991;**40**:1007–1012.
104. Sassy-Prigent C, Heudes D, Mandet C, et al. Early glomerular macrophage recruitment in streptozotocin-induced diabetic rats. *Diabetes* 2000;**49**:466–475.
105. Navarro JF, Milena FJ, Mora C, Leon C, Garcia J. Renal pro-inflammatory cytokine gene expression in diabetic nephropathy: Effect of angiotensin-converting enzyme inhibition and pentoxifylline administration. *Am J Nephrol* 2006;**26**:562–570.
106. Sekizuka K, Tomino Y, Sei C, et al. Detection of serum IL-6 in patients with diabetic nephropathy. *Nephron* 1994;**68**:284–285.
107. Shikano M, Sobajima H, Yoshikawa H, et al. Usefulness of a highly sensitive urinary and serum IL-6 assay in patients with diabetic nephropathy. *Nephron* 2000;**85**:81–85.
108. Wong CK, Ho AW, Tong PC, et al. Aberrant activation profile of cytokines and mitogen-activated protein kinases in type 2 diabetic patients with nephropathy. *Clin Exp Immuno* 2007;**149**:123–131.
109. Moriwaki Y, Yamamoto T, Shibutani Y, et al. Elevated levels of interleukin-18 and tumor necrosis factor-alpha in serum of patients with type 2 diabetes mellitus: Relationship with diabetic nephropathy. *Metabolism* 2003;**52**:605–608.
110. Pickup JC, Chusney GD, Thomas SM, Burt D. Plasma interleukin-6, tumour necrosis factor alpha and blood cytokine production in type 2 diabetes. *Life Sci* 2000;**67**:291–300.
111. Dalla VM, Mussap M, Gallina P, et al. Acute-phase markers of inflammation and glomerular structure in patients with type 2 diabetes. *J Am Soc Nephrol* 2005;**16**(Suppl 1):S78–S82.
112. Suzuki D, Miyazaki M, Naka R, et al. In situ hybridization of interleukin 6 in diabetic nephropathy. *Diabetes* 1995;**44**:1233–1238.
113. Melnikov VY, Ecdet T, Fantuzzi G, et al. Impaired IL-18 processing protects caspase-1-deficient mice from ischemic acute renal failure. *J Clin Invest* 2001;**107**:1145–1152.
114. Miyauchi K, Takiyama Y, Honjyo J, Tateno M, Haneda M. Upregulated IL-18 expression in type 2 diabetic subjects with nephropathy: TGF-beta1 enhanced IL-18 expression in human renal proximal tubular epithelial cells. *Diabetes Res Clin Pract* 2009;**83**:190–199.
115. Altinova AE, Yetkin I, Akbay E, Bukan N, Arslan M. Serum IL-18 levels in patients with type 1 diabetes: Relations to metabolic control and microvascular complications. *Cytokine* 2008;**42**:217–221.
116. Nakamura A, Shikata K, Hiramatsu M, et al. Serum interleukin-18 levels are associated with nephropathy and atherosclerosis in Japanese patients with type 2 diabetes. *Diabetes care* 2005;**28**:2890–2895.
117. Araki S, Haneda M, Koya D, et al. Predictive impact of elevated serum level of IL-18 for early renal dysfunction in type 2 diabetes: An observational follow-up study. *Diabetologia* 2007;**50**:867–873.
118. Esposito K, Nappo F, Marfella R, et al. Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans: Role of oxidative stress. *Circulation* 2002;**106**:2067–2072.
119. Arnalich F, Hernandez A, Lopez-Maderuelo D, et al. Enhanced acute-phase response and oxidative stress in older adults with type II diabetes. *Horm Metab Res* 2000;**32**:407–412.
120. Schmidt AM, Yan SD, Wautier JL, Stern D. Activation of receptor for advanced glycation end products: A mechanism for chronic vascular dysfunction in diabetic vasculopathy and atherosclerosis. *Circ Res* 1999;**84**:489–497.
121. Miyata T, Dan T. Inhibition of advanced glycation end products (AGEs): An implicit goal in clinical medicine for the treatment of diabetic nephropathy? *Diabetes Res Clin Pract* 2008;**82**(Suppl 1):S25–S29.

122. Ogawa S, Mori T, Nako K, Kato T, Takeuchi K, Ito S. Angiotensin II type 1 receptor blockers reduce urinary oxidative stress markers in hypertensive diabetic nephropathy. *Hypertension* 2006;**47**:699–705.
123. Sugimoto H, Shikata K, Wada J, Horiuchi S, Makino H. Advanced glycation end products-cytokine-nitric oxide sequence pathway in the development of diabetic nephropathy: Aminoguanidine ameliorates the overexpression of tumour necrosis factor- $\alpha$  and inducible nitric oxide synthase in diabetic rat glomeruli. *Diabetologia* 1999;**42**:878–886.
124. Pamir N, McMillen TS, Kaiyala KJ, Schwartz MW, LeBoeuf RC. Receptors for tumor necrosis factor- $\alpha$  play a protective role against obesity and alter adipose tissue macrophage status. *Endocrinology* 2009;**50**:4124–4134.
125. Luo SF, Fang RY, Hsieh HL, et al. Involvement of MAPKs and NF- $\kappa$ B in tumor necrosis factor  $\alpha$ -induced vascular cell adhesion molecule 1 expression in human rheumatoid arthritis synovial fibroblasts. *Arthritis Rheum* 2009;**62**:105–116.
126. O'Hara AM, Bhattacharyya A, Bai J, Mifflin RC, Ernst PB, Mitra S, Crowe SE. Tumor necrosis factor (TNF)- $\alpha$ -induced IL-8 expression in gastric epithelial cells: Role of reactive oxygen species and AP endonuclease-1/redox factor (Ref)-1. *Cytokine* 2009;**46**:359–369.
127. Cao YL, Wang YX, Wang DF, Meng X, Zhang J. Correlation between omental TNF- $\alpha$  protein and plasma PAI-1 in obesity subjects. *Int J Cardiol* 2008;**128**:399–405.
128. Romanatto T, Roman EA, Arruda AP, et al. Deletion of tumor necrosis factor- $\alpha$  receptor 1 (TNFR1) protects against diet-induced obesity by means of increased thermogenesis. *J Biol Chem* 2009;**284**:36213–36222.
129. Kaddai V, Jager J, Gonzalez T, et al. Involvement of TNF- $\alpha$  in abnormal adipocyte and muscle sortilin expression in obese mice and humans. *Diabetologia* 2009;**52**:932–940.
130. McCarthy ET, Sharma R, Sharma M, et al. TNF- $\alpha$  increases albumin permeability of isolated rat glomeruli through the generation of superoxide. *J Am Soc Nephrol* 1998;**9**:433–438.
131. Javaid B, Quigg RJ. Treatment of glomerulonephritis: Will we ever have options other than steroids and cytotoxics? *Kidney Int* 2005;**67**:1692–1703.
132. Aringer M, Smolen JS. The role of tumor necrosis factor- $\alpha$  in systemic lupus erythematosus. *Arthritis Res Ther* 2008;**10**:1–8.
133. Feldmann M, Pusey CD. Is there a role for TNF- $\alpha$  in anti-neutrophil cytoplasmic antibody-associated vasculitis? Lessons from other chronic inflammatory diseases. *J Am Soc Nephrol* 2006;**17**:1243–1252.
134. Navarro JF, Milena FJ, Mora C, et al. Tumor necrosis factor- $\alpha$  gene expression in diabetic nephropathy: Relationship with urinary albumin excretion and effect of angiotensin-converting enzyme inhibition. *Kidney Int Suppl* 2005;**99**:S98–S102.
135. Kalantarinia K, Awad AS, Siragy HM. Urinary and renal interstitial concentrations of TNF- $\alpha$  increase prior to the rise in albuminuria in diabetic rats. *Kidney Int* 2003;**64**:1208–1213.
136. Navarro JF, Mora C, Muros M, Garcia J. Urinary tumour necrosis factor- $\alpha$  excretion independently correlates with clinical markers of glomerular and tubulointerstitial injury in type 2 diabetic patients. *Nephrol Dial Transplant* 2006;**21**:3428–3434.
137. DiPetrillo K, Coutermarsh B, Gesek FA. Urinary tumor necrosis factor contributes to sodium retention and renal hypertrophy during diabetes. *Am J Physiol Renal Physiol* 2003;**284**:F113–F121.
138. Yoshida LS, Tsunawaki S. Expression of NADPH oxidases and enhanced H(2)O(2)-generating activity in human coronary artery endothelial cells upon induction with tumor necrosis factor- $\alpha$ . *Int Immunopharmacol* 2008;**8**:1377–1385.
139. Zhou Z, Wang L, Song Z, Lambert JC, McClain CJ, Kang YJ. A critical involvement of oxidative stress in acute alcohol-induced hepatic TNF- $\alpha$  production. *Am J Pathol* 2003;**163**:1137–1146.
140. Kuhad A, Chopra K. Attenuation of diabetic nephropathy by tocotrienol: Involvement of NF $\kappa$ B signaling pathway. *Life Sci* 2009;**84**:296–301.
141. Ebenezer PJ, Mariappan N, Elks CM, Haque M, Francis J. Diet-induced renal changes in Zucker rats are ameliorated by the superoxide dismutase mimetic TEMPOL. *Obesity* 2009;**17**:1994–2002.
142. Niu J, Kolattukudy PE. Role of MCP-1 in cardiovascular disease: Molecular mechanisms and clinical implications. *Clin Sci* 2009;**117**:95–109. Review.
143. Mehrabian M, Sparkes RS, Mohandas T, Fogelman AM, Lusic AJ. Localization of monocyte chemoattractant protein-1 gene (SCYA2) to human chromosome 17q11.2-q21.1. *Genomics* 1991;**9**:200–203.
144. Craig MJ, Loberg RD. CCL2 (monocyte chemoattractant protein-1) in cancer bone metastases. *Cancer Metastasis Rev* 2006;**25**:611–619.
145. Carr MW, Roth SJ, Luther E, Rose SS, Springer TA. Monocyte chemoattractant protein 1 acts as a T-lymphocyte chemoattractant. *Proc Natl Acad Sci U S A* 1994;**91**:3652–3656.
146. Chow FY, Nikolic-Paterson DJ, Ozols E, Atkins RC, Rollin BJ, Tesch GH. Monocyte chemoattractant protein-1 promotes the development of diabetic renal injury in streptozotocin-treated mice. *Kidney Int* 2006;**69**:73–80.
147. Ha H, Yu MR, Choi YJ, Kitamura M, Lee HB. Role of high glucose-induced nuclear factor- $\kappa$ B activation in monocyte chemoattractant protein-1 expression by mesangial cells. *J Am Soc Nephrol* 2002;**13**:894–902.
148. Yamagishi S, Inagaki Y, Okamoto T, et al. Advanced glycation end product-induced apoptosis and overexpression of vascular endothelial growth factor and monocyte chemoattractant protein-1 in human-cultured mesangial cells. *J Biol Chem* 2002;**277**:20309–20315.
149. Gu L, Hagiwara S, Fan Q, et al. Role of receptor for advanced glycation end-products and signalling events in advanced glycation end-product-induced monocyte chemoattractant protein-1 expression in differentiated mouse podocytes. *Nephrol Dial Transplant* 2006;**21**:299–313.
150. Wu YG, Lin H, Qi XM, et al. Prevention of early renal injury by mycophenolate mofetil and its mechanism in experimental diabetes. *Int Immunopharmacol* 2006;**6**:445–453.
151. Wu YG, Lin H, Qian H, Zhao M, Qi XM, Wu GZ, Lin ST. Renoprotective effects of combination of angiotensin converting enzyme inhibitor with mycophenolate mofetil in diabetic rats. *Inflamm Res* 2006;**55**:192–199.
152. Tesch GH. MCP-1/CCL2: A new diagnostic marker and therapeutic target for progressive renal injury in diabetic nephropathy. *Am J Physiol Renal Physiol* 2008;**294**:F697–F701.
153. Morii T, Fujita H, Narita T, et al. Association of monocyte chemoattractant protein-1 with renal tubular damage in diabetic nephropathy. *J Diabetes Complications* 2003;**17**:11–15.
154. Mezzano S, Aros C, Droguett A, et al. NF- $\kappa$ B activation and overexpression of regulated genes in human diabetic nephropathy. *Nephrol Dial Transplant* 2004;**19**:2505–2512.
155. Banba N, Nakamura T, Matsumura M, Kuroda H, Hattori Y, Kasai K. Possible relationship of monocyte chemoattractant protein-1 with diabetic nephropathy. *Kidney Int* 2000;**58**:684–690.
156. Wu Y, Wu G, Qi X, Lin H, Qian H, Shen J, Lin S. Protein kinase C beta inhibitor LY333531 attenuates intercellular adhesion molecule-1 and monocyte chemoattractant protein-1 expression in the kidney in diabetic rats. *J Pharmacol Sci* 2006;**101**:335–343.
157. Chiarelli F, Cipollone F, Mohn A, et al. Circulating monocyte chemoattractant protein-1 and early development of nephropathy in type 1 diabetes. *Diabetes Care* 2002;**25**:1829–1834.
158. Cipollone F, Chiarelli F, Iezzi A, et al. Relationship between reduced BCL-2 expression in circulating mononuclear cells and early nephropathy in type 1 diabetes. *Int J Immunopathol Pharmacol* 2005;**18**:625–635.
159. Heidland A, Sebekova K, Schinzel R. Advanced glycation end products and the progressive course of renal disease. *Am J Kidney* 2001;**38**(Suppl 1):S100–S106. Review.
160. Koga M, Otsuki M, Kubo M, Hashimoto J, Kasayama S. Relationship between circulating vascular cell adhesion molecule-1 and microvascular complications in type 2 diabetes mellitus. *Diabet Med* 1998;**15**:661–667.
161. Ina K, Kitamura H, Okeda T, Nagai K, Liu ZY, Matsuda M, Fujikura Y. Vascular cell adhesion molecule-1 expression in the renal interstitium of diabetic KKAY mice. *Diabetes Res Clin Pract* 1999;**44**:1–8.
162. Murakami H, Tamasawa N, Matsui J, Yamato K, JingZhi G, Suda T. Plasma levels of soluble vascular adhesion molecule-1 and cholesterol oxidation product in type 2 diabetic patients with nephropathy. *J Atheroscler Thromb* 2001;**8**:21–24.
163. Jenkins AJ, Zhang SX, Rowley KG, et al. Increased serum pigment epithelium-derived factor is associated with microvascular complications,

- vascular stiffness and inflammation in Type 1 diabetes. *Diabet Med* 2007;**24**:1345–1351.
164. Okada S, Shikata K, Matsuda M, et al. Intercellular adhesion molecule-1-deficient mice are resistant against renal injury after induction of diabetes. *Diabetes* 2003;**52**:2586–2593.
  165. Lane TA, Lamkin GE, Wancewicz E. Modulation of endothelial cell expression of intercellular adhesion molecule 1 by protein kinase C activation. *Biochem Biophys Res Commun* 1989;**161**:945–952.
  166. Hirata K, Shikata K, Matsuda M, Akiyama K, Sugimoto H, Kushiro M, Makino H. Increased expression of selectins in kidneys of patients with diabetic nephropathy. *Diabetologia* 1998;**41**:185–192.
  167. Leone M, Boutière-Albanèse B, Valette S, et al. Cell adhesion molecules as a marker reflecting the reduction of endothelial activation induced by glucocorticoids. *Shock* 2004;**21**:311–314.
  168. Matsui H, Suzuki M, Tsukuda R, Iida K, Miyasaka M, Ikeda H. Expression of ICAM-1 on glomeruli is associated with progression of diabetic nephropathy in a genetically obese diabetic rat, Wistar fatty. *Diabetes Res Clin Pract* 1996;**32**:1–9.
  169. Booth G, Stalker TJ, Lefer AM, Scalia R. Mechanisms of amelioration of glucose-induced endothelial dysfunction following inhibition of protein kinase C in vivo. *Diabetes* 2002;**51**:1556–1564.
  170. Coimbra TM, Janssen U, Grone HJ, et al. Early events leading to renal injury in obese Zucker (fatty) rats with type II diabetes. *Kidney Int* 2000;**57**:167–182.
  171. Lavaud S, Michel O, Sassy-Prigent C, et al. Early influx of glomerular macrophages precedes glomerulosclerosis in the obese Zucker rat model. *J Am Soc Nephrol* 1996;**7**:2604–2615.
  172. Sugimoto H, Shikata K, Hirata K, et al. Increased expression of intercellular adhesion molecule-1 (ICAM-1) in diabetic rat glomeruli: Glomerular hyperfiltration is a potential mechanism of ICAM-1 upregulation. *Diabetes* 1997;**46**:2075–2081.
  173. Chow FY, Nikolic-Paterson DJ, Ozols E, Atkins RC, Tesch GH. Intercellular adhesion molecule-1 deficiency is protective against nephropathy in type 2 diabetic db/db mice. *J Am Soc Nephrol* 2005;**16**:1711–1722.
  174. Ye SD, Zheng M, Zhao LL, et al. Intensive insulin therapy decreases urinary MCP-1 and ICAM-1 excretions in incipient diabetic nephropathy. *Eur J Clin Invest* 2009;**39**:980–985.
  175. Rubio-Guerra AF, Vargas-Robles H, Lozano Nuevo JJ, Escalante-Acosta BA. Correlation between circulating adhesion molecule levels and albuminuria in Type-2 diabetic hypertensive patients. *Kidney Blood Press Res* 2009;**32**:106–109.
  176. Clausen P, Jacobsen P, Rossing K, Jensen JS, Parving HH, Feldt-Rasmussen B. Plasma concentrations of VCAM-1 and ICAM-1 are elevated in patients with Type 1 diabetes mellitus with microalbuminuria and overt nephropathy. *Diabet Med* 2000;**17**:644–649.
  177. Kosugi T, Nakayama T, Heinig M, et al. Effect of lowering uric acid on renal disease in the type 2 diabetic db/db mice. *Am J Physiol Renal Physiol* 2009;**297**:F481–F488.
  178. Onozato ML, Tojo A, Goto A, Fujita T. Radical scavenging effect of gliclazide in diabetic rats fed with a high cholesterol diet. *Kidney Int* 2004;**65**:951–960.
  179. Wang L, Zhang L, Yu Y, Wang Y, Niu N. The protective effects of taurine against early renal injury in STZ-induced diabetic rats, correlated with inhibition of renal LOX-1-mediated ICAM-1 expression. *Ren Fail* 2008;**30**:763–771.